Long-term Cell Tracer

Name:	Long-term Cell Tracer 580 Yellow, Cat.# P710Y (1 ml), P710YS (0.1 ml)			
name:	Long-term Cell Tracer 500 Green, Cat.# P710G (1 ml), P710GS (0.1 ml)			
Application:	Cell Tracer is for long-term tracing of a wide range of cell types, including cancer cells, bone marrow stromal cell (BMSC), peripheral blood mononuclear cell, endothelial progenitor cell, human/mouse mesenchymal stem cells, skin stem cells <i>et al.</i> This product is for research use only.			
		Excitation	Emission	
Excitation / Emission:	Cell Tracer 580	405nm or 488nm	580nm (550nm ~ 600nm)	
	Cell Tracer 500	405nm	500nm (450nm ~ 525nm)	
Photostability:	<i>in vitro</i> tracing: 12 generations <i>in vivo</i> tracing: 3 weeks			
Product Description:	Cell Tracers are a class of organic fluorescent dots with comparable size a photostability to inorganic quantum dots (QDs) to overcome the limitations quantum dots (e.g., potential toxicity and compromised fluorescence in presence ROS) in advanced bio-imaging applications. Upon conjugation with a penetrating peptide, Cell Tracer shows excellent labeling efficiency to living co and outperforms the current gold standard inorganic quantum dots cell label reagents, in long term <i>in vitro / in vivo</i> cell tracing (Fig. 1 & Table 1.). Stem cell tracing study also suggests that Cell Tracer has no negative effect mesenchymal stem cell differentiation (Table 2). The merits of Cell Tracer mathem promising alternatives to quantum dot probes, which is of high important.			
	for translational research applications. Additionally, the biocompatible polymeric matrix endows this kind of organic fluorescent dots customized surface functional groups for further modification/conjugation with a variety of biomolecules for specific imaging tasks besides cell tracking (Table 2).			
	 Long term photostability - 3 weeks <i>in vivo</i> High brightness - high signal to noise ratio Different colors - blue, green, yellow, red Low toxicity - no heavy metal elements Customization friendly - can conjugate with desired biomolecules easy to use - only one component 			
Shipping / Storage:	Ship at RT. Store at 4	•°C, — 20°C or — 80°C (A	woid repeated freeze-thaw cycles.)	

Shelf Life:	1 months at RT, 3 mon	1 months at RT, 3 months at 4° C, 12 months at -20° C or -80° C (preferred).			
Component:	Cell Tracer 580	P710Y	1 mL		
		P710YS	0.1 mL		
		P710G	1 mL		
	Cell Tracer 500	P710GS	0.1 mL		
	Concentration: 200 nN	Concentration: 200 nM			
Remark:		The different colors of Cell Tracers allow simultaneously tracing of two different groups of cells to investigate their migration and interaction.			

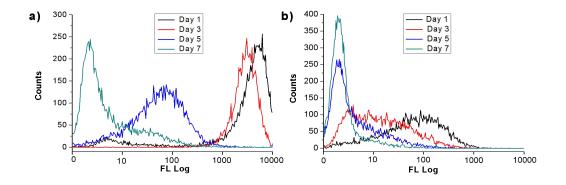


Figure 1.

Flow cytometry overlay histograms of MCF-7 cells at different time point after labeling with (**a**) 2 nM Cell Tracer or (**b**) 2 nM quantum dot.

	Day 1	Day 3	Day 5	Day 7
Cell Tracer	99.4%	98.2%	82.2%	31.1%
Other QDs	84.1%	43.9%	26.9%	4.3%

Table 1 summarizes the fluorescence intensity of the labeled cells at different time point from flow cytometry data in Fig. 1. These data show that Cell Tracer last much longer in labeled cells than other QDs.

Table 2. Comparison of Cell Tracer and other QDs

	Working concentration	Low Toxicity	Negative effect on stem cells	Customized targeting ability	Tracing ability
Cell Tracer	0.1 - 2 nM	v	V	V	9-12 generations
Other QDs	2 - 15 nM	×	×	×	5-6 generations

Cell Tracer has advantage over other QDs in many aspects including working concentration, toxicity and flexibility etc.

Protocol

The optimal working concentration of the Cell Tracer is typically in the range of 0.1 nM to 4 nM depending on the cell type and application. We recommend to test serial dilution test to figure the optimized staining condition for your cells. The following protocols use 2 nM Cell Tracer as example.

Make Cell Labeling Medium

Add 10 μ L Cell Tracer (200 nM) to 1 mL complete cell culture medium, vortex for 30 seconds. Now the **Cell Labeling medium** containing 2 nM Cell Tracer is ready to use.

Always prepare the labeling medium freshly.

Labeling Adherent Cells (example of labeling in 6-well plate)

Plating cells:

Seed the cells in desired culture dish / flask. Cell density may vary depending on the cell type. Cells can be cultured on coverslip for special assay. The cells can be labeled when they attach and reach ~80% confluency. The time window could be a few hours to overnight.

Labeling:

- 1. Wash the cells twice using PBS.
- 2. Add 1 mL cell labeling medium into each well and incubate at 37 °C for 4 h to overnight.
- 3. Wash the cells twice with PBS.

Optional: If desired, the labeled cells can be fixed at this point. Wash the cells 3 times with PBS, and then fix with 3.7% formaldehyde in PBS for 15 minutes at room temperature. Wash 3 times post-fixation in PBS prior to imaging.

4. The labeled cells are ready for further *in vivo* or *in vitro* assay.

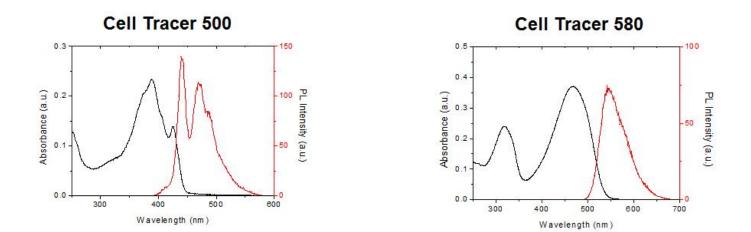
Labeling non-Adherent Cells

- 1. Collect the cells and centrifuge at 1500 rpm for 5 min. Discard the medium.
- 2. Add cell labeling medium to resuspend the cell pallet at 1×10^{6} cells / 3 mL medium
- 3. Incubate at 37 °C for 4 hours.
- 4. Centrifuge at 1500 rpm for 5 min and wash the cells twice with PBS.

Optional: If desired, the labeled cells can be fixed at this point.

5. The labeled cells are ready for further *in vivo* or *in vitro* assay.

Fluorescence Spectrum:



Confocal imaging parameters:

If used separately

Cell Tracer 500 (Green): 405 nm excitation, 420-600 nm any bandpass or long pass filters.

Cell Tracer 580 (Yellow): 488 nm excitation, 505 nm above long pass filter.

If use the two probes for two groups of cells to simultaneously perform dual-color cell tracking:

Cell Tracer 500 (Green): 405 nm excitation, 420-480 nm / 420-450 nm bandpass

Cell Tracer 580 (Yellow): 488 nm/514 nm excitation, 535-565 nm bandpass/560 nm above long pass filter.

Carefully tune the confocal parameters, such as gain, laser power and bandpass filters, it will give two distinct signals from green and yellow dots without crosstalking.

Note: The availability of filters depends on confocal microscopies, the operator always can try and find the optimized filters to obtain maximized signal.

-- The end –